

Journal of Experimental Biology and Agricultural Sciences

http://www.jebas.org

ISSN No. 2320 - 8694

DEVELOPMENT OF A PROTOCOL FOR THE APPLICATION OF COMMERCIAL BIO-STIMULANT MANUFACTURED FROM *Kappaphycus alvarezii* IN SELECTED VEGETABLE CROPS

Kosalaraman Karthikeyan and Munisamy Shanmugam*

Research and Development Division, AquAgri Processing Private Limited, B5, SIPCOT Industrial Complex, Manamadurai - 630 606. Sivaganga District, Tamil Nadu, INDIA

Received – January 12, 2016; Revision – January 27, 2016; Accepted – February 20, 2016 Available Online – February 20, 2016

DOI: http://dx.doi.org/10.18006/2016.4(1).92.102

KEYWORDS

Application protocol

Seaweed extract

Kappaphycus alvarezii

Vegetable crops

Yield and quality

ABSTRACT

The field study was conducted to develop a protocol for application of commercially manufactured bio-stimulant (Brand name: AquaSap) from seaweed *Kappaphycus alvarezii*. Efficacy of the bio-stimulant was tested at 5% through foliar application in selected important vegetable crops. 3 to 4 applications were applied based on the crop cycle of the plant. Total 27 vegetable crops were studied during 2012 to 2015 and observed their response towards bio-stimulant applied in terms of general health of the plant, growth, yield and quality of the vegetable produce. 11% to 52% of yield increases were observed with improved quality in all 27 crops studied. Therefore seaweed bio-stimulants will have enormous potential to organic vegetable production in future.

E-mail: m.shanmugam@aquagri.in (Muniyasamy Shanmugam)

Peer review under responsibility of Journal of Experimental Biology and Agricultural Sciences.

Production and Hosting by Horizon Publisher (http://publisher.jebas.org/index.html).
All rights reserved.

All the article published by Journal of Experimental Biology and Agricultural Sciences is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License Based on a work at www.jebas.org.



^{*} Corresponding author

93 Karthikeyan and Shanmugam

1 Introduction

Seaweed personifies not only as an alternative to conventional chemical fertilizers but also chronically used in agriculture, horticulture, cookies, ice-cream and jelly mix (Sumkiman et al., 2014). Further, it was well reported that seaweed extract contain nutrient of major and minor element, vital amino acid, essential vitamins and plant growth regulators which stimulate the growth and quality yield of crops. Application of seaweed liquid extract stimulate different aspects of plant like good health, development of root system, absorption of mineral, enlargement of shoot, increased rate of photosynthesis and crop yield (Sridhar & Rengasamy, 2010). Seaweed liquid extract have newly gained importance as foliar spray for lots of crops including various variety of grasses, flowers, cereals, vegetables and spices (Pramanick et al. 2013 & 2014). Further, Zodape (2001) tried various modes of seaweed extract application such as a foliar spray, application to soil and soaking of seeds before sowing and reported that extract not only enhances the germination of seeds but also increases uptake of plant nutrients and gives resistance to frost and fungal diseases.

The aqueous extracts of the alga *Codium fragile* was effective in increasing root length and it is 18.0% longer than the control in soybeans (Anisimov & Chaikina, 2014). Furthermore, Pise & Sabale (2010) treated fenugreek with 50% of seaweed and reported improvement in the concentration of carbohydrate, proteins, free amino acids, polyphenols and nitrogen content while comparing with control plants. Similarly, yield and nutrient content value were found higher in banana when treated with 5% of bio-stimulant (AQUASAP) of *Kappaphycus alvarezii* (Karthikeyan & Shanmugam, 2014). Vegetables are herbaceous plants and produce large amount of biomass within short period (Chatterjee & Thirumdasu, 2014). Vegetables are very essential to human health as they are rich in dietary fibre and source of essential vitamins, minerals, trace elements, vitamins and antioxidants.

In India, vegetable production was around 146.55 million tons from an area of 8.5 million hectare during 2010-2011. The 4 major vegetables viz. potato (28.9%), tomato (11.3%), onion (10.3%) and brinjal (8.1%) contribute 58.6% of total vegetable production. Other important vegetables are cabbage (5.4%), cauliflower (4.6%), okra (3.9%), peas (2.4%) and okra contribute 73% of total world production (Vanitha et al., The bio-stimulant manure from red seaweed K. alvarezii is well-off in potash with other primary nutrients like N, P, K and secondary nutrients like Cu, Zn, Fe, Mo, Mn, etc., in addition and to significant amount of plant growth regulators (Zodape et al., 2009; Prasad et al., 2010; Karthikeyan & Shanmugam, 2014). The present investigation describes the dosage and application protocol of bio-stimulant manufactured from K. alvarezii (AQUASAP is brand name of AquAgri) on some selected 27 vegetables crops for yield and quality improvement.

2 Materials and Methods

The trial was carried out at R&D plot of AquAgri Processing Private Limited and in the farmers' field in Manamadurai, Sivagangai Dt., Tamil Nadu, India. (Latitude is 9°42′56′N and longitude 78°28′2′E). The annual normal rainfall received by the district is 850 mm. The experiment trial was conducted in 8 plots with 6 m x 4m for each vegetable crop studied. The healthy seeds were selected and sowed carefully into the field and the trial crops were irrigated periodically and chemical fertilizers were applied to crops as per the recommendation of National Horticulture Board, India. Biostimulant (Aquasap) was collected from the stock of AquAgri Processing Private Limited and 5% solution was prepared and

2.1 Application protocol of bio-stimulant Aquasap for vegetable crops

Bio-stimulant (Brand name: AquaSap) manufactured from *K. alvarezii* was applied to the crops tested in the present investigation through foliar application. Three doses viz. vegetative, pre-flowering and post flowering stages were given to short-term plants whereas four doses were applied to long-term crops. Table 2-6 shows the application protocol for 27 crops tested in this study. The physico-chemical and nutritive value of the bio-stimulant (AquaSap) has been given in table 1.

2.1.1 Tomato (Solanum lycopersicum L.)

The trial on tomato (Co3 hybrid) was conducted in June 2012 (Table 2.) The seeds were sowed in nursery beds, then nursery plants were collected after 25th day of sowing and their roots were dipped at 0.7% of bio-stimulant Aquasap for 10min before transplantation. The first spray was given on 10th day of transplantation, second and third doses were sprayed on 25-30d (pre-flowering stage) and on 45-50d (flowering stage) respectively and last dose was applied at first picking stage (Table 3).

2.1.2 Lady's finger (Okra) (Abelmoschus esculentus (L) Moench) and Brinjal (Solanum melongeana L.)

The experiment of lady's finger (var. US 7902) and brinjal Co2 hybrid was also conducted in 2012 (Table 2). The okra seeds were soaked in 1% of bio-stimulant for 10 min. and the soaked seeds were sowed into the field directly. Treated seeds were also sowed in nursery beds and nursery plants (35d old) were collected, treated their roots with 0.7% of bio-stimulant for 10 min before transplantation. The application of bio-stimulant through foliar was given at the vegetative stage (15-20d), second spray at flowering stage (35-40d) and final spray was at first fruits picking stage (50-55d) (Table 3).

Table 1 Physico-chemical properties and Nutritive status of Aquasap bio-stimulant from seaweed K. alvarezii.

Parameters	Units	Results	Parameters	Units	Results
Physical properties of AquaSap			Nutritive Value (Amino Acid)		
			Alanine	g/100g	0.014
Organic Matter (%)	gm/100g	0.65	Arginine	g/100g	0.0003
Specific Gravity	-	1.14	Aspartic acid	g/100g	0.0019
Electrical Conductivity	dSm ⁻¹	63.3	Cystine	g/100g	0.0017
pH (1% solution)	-	6.68	Glycine	g/100g	0.065
Moisture content (%)	gm/100g	94.82	Histidine	g/100g	0.0007
Total Ash (%)	gm/100g	4.53	Isoleucine	g/100g	0.0022
Macro and Micro	Nutrient contents Aqua	aSap	Leucine	g/100g	0.0022
			Lysine	g/100g	0.019
Parameters	Units	Results	Tryptophan	g/100g	0.007
Nitrogen (N)	gm/100g	0.007	Methionine	g/100g	0.0007
Phosphorous (P)	mg/kg	3.57	Phenylalanine	g/100g	0.0028
Potash (K)	gm/100g	1.50	Proline	g/100g	0.053
Sodium (Na)	gm/100g	0.26	Serine	g/100g	0.0013
Calcium (Ca)	gm/100g	0.03	Threonine	g/100g	0.0006
Silica (Si)	gm/100g	0.02	Tyrosine	g/100g	0.0016
Chlorine (Cl)	gm/100g	2.15	Valine	g/100g	0.0026
Magnesium (Mg)	mg/kg	0.04	Glutamic acid	g/100g	0.0022
Iron (Fe)	gm/100g	16.95	Nutriti	ve Value (Vitami	ns)
Sulphur (S)	gm/100g	0.03	Vitamin - A	IU/100g	3363.44
Boron as (B)	mg/kg	768	Vitamin – E	IU/100g	0.21
Copper (Cu)	mg/kg	1.1	Vitamin – C	mg/100g	22.52
Zinc (Zn)	mg/kg	2.15	Vitamin – B1	mg/100g	0.007
Manganese (Mn)	mg/kg	5.93	Vitamin –B5	mg/100g	301.1
Cobalt (Co)	mg/kg	0.92	Vitamin – B6	mg/100g	3170.2

2.1.3 Chillies (Capsicum annuum L. Var. annuum)

Hybrid chilli (US 612) was selected for present study and its seeds were sowed directly into field. The seaweed biostimulant was applied at vegetative stage (40-45d), at flowering stage (90-100d) and last dose was given at first fruits picking stage (125-130d). In the case of transplanted plant, nurseries were created (40d old) and treated their roots at 0.7% of bio-stimulant for 10min before transplantation. During growing period, first dose of bio-stimulant was given at 20-25th day of transplantation and second and last applications were given at 60-65th day (i.e. flowering stage) and at 80-85th day of transplantation (Table 3) respectively.

2.1.4 Capsicum (Capsicum annuum L)

Trial on capsicum (var. Arka Mohini) was carried out in January 2014 (Table 2). The bio-stimulant AquaSap was applied at vegetative (30-35d), flowering stage (60-65d) and fruits picking stage (90-95d). But in the case of transplants raised from 40d old nurseries whose roots were treated with 0.7% of bio-stimulant for 10min before transplantation, first dose was given at 20-25d (vegetative stage), 60-65d (flowering

stage) and at 80-85d (first fruits picking stage) day of transplantation (Table 3).

2.1.5 Variety of Gourds

The experimental study on nine varieties of gourds, i.e., Ash gourd (var. MAH-1), Pumpkin (Arka Chandan), Snake gourd (Covai -951), Ridge gourd (US 66), Bottle gourd (WARAD MGH-4), Bitter gourd (US 475), Cucumber (local variety), Watermelon (Ankur Kashish) and Chow chow (Green Fruits) (Table 2) was conducted during 2012 to 2014. The seeds were soaked in 1% of bio-stimulant for 30 min. and the seeds were sowed in the study field. The application of bio-stimulant was given at vegetative stage (20-25d), flowering stage (60-65d) and first fruits picking stage (80-85d). In the case of chow chow, mature fruits were planted in the field, and bio-stimulant was first applied at vegetative phase of 25-30 days of plantation, pre-flowering phase (3rd month) and final dose was given at flowering phase (5th month) (Table 3). In the cucumber, first spray was done at germination phase (10-15th day), followed by second spray at 35-40th day (vegetative stage) and final dose was applied at flowering initiation stage (65-70d) (Table 4).

2.1.6 Cole crops, Root and Tuber vegetables

2.1.6.1 Potato (Solanum tuberosum L.)

The potato (var. Kufri Jyoti) trial was conducted in August 2013 (Table 2). Bio-stimulant AquaSap was applied at plant establishment stage (20-25d) vegetative phase (50-55d), early root development stage (80-85d) and last dosage was given at maturity stage (100-105d) (Table 5).

2.1.6.2 Cabbage (Brassica oleracea var. capitata L)

Maharani- F1, a hybrid variety of cabbage was taken for trial in January 2013 (Table 2). 40 days old nursery plant was created and root treatment was given at 0.7% of bio-stimulant for 10min before transplantation. Field application of bio-stimulant was applied at plant establishment stage (10-15d), second dose was sprayed at head initiation stage (35-40d) and last spray was sprayed at head development phase (70-75d) (Table 5).

2.1.6.3 Cauliflower (Brassica oleracea var. botrytis)

In January 2013, cauliflower (var. Shobha F1) was taken for trial (Table 2). The root of nursery plants (35d) raised was treated with 0.7% of bio-stimulant for 10min and transplanted. During crop cycle, first spray of bio-stimulant aquasap was given at plant establishment stage (10-15d), the second dose at curd initiation stage (25-30d) and last dose was given at curd development stage (45-50d) (Table 5).

2.1.6.4 Beetroot (*Beta vulgaris* L) and Carrot (*Daucus carota* L)

The studies on beetroot (Vally Queen) as well as carrot (Pusa Kesar) were conducted in 2013 and bio-stimulant Aquasap was applied at vegetative stage (25-30d), early root development stage (55-60d) and root maturity stage (80 -85d) (Table 5).

2.1.6.5 Radish (Raphanus sativus L) and Knol-Khol (Brassica caulorapa)

Radish (Roshni) and knol-khol (Early White) trial was undertaken in January 2013. The bio-stimulant was given at 10-15th (vegetative stage), 25-30th (early root development stage) and at 40 - 45th day of sowing (root maturity stage) (Table 5).

2.1.7 Other vegetable crops

2.1.7.1 Lima Bean (*Phaseolus lunatus* L) and Dolichos Bean (*Lab lab purpureus* var. *typicus*)

The experiment on Lima (Co2) and Dolichos (Ankur Goldy) beans were conducted during 2012. The seeds were soaked in 1% of bio-stimulant for 10min then sowed into the field. During crop period, three spray of bio-stimulant were given viz. at vegetative phase (20-25d), flowering stage (40-45d),

pod formation stage (60-65d) and last spray was given at first picking stage (80-85th day of sowing) (Table 6).

2.1.7.2 Soybean (Glycine max (L.) Marr.)

The experiment on soybean (JSS 355) was conducted in July, 2012. The seeds were soaked in 1% of bio-stimulant for 10min and then the seeds were carefully sowed in the field. The crop was applied with bio-stimulant Aquasap for four times viz. at 20-25d, 40-45d, 60-65d and at 80-85th day of sowing (Table 6).

2.1.7.3 Moringa (Moringa oleifera L.)

The efficacy trial of AquaSap on drumstick (PKM-1) was conducted in 2012 (Table 2). The seeds were soaked in 1% of bio-stimulant for 10min and during crop period bio-stimulant aquasap was applied at nurseries stage (25-30th), pre-flowering phase (3rd month), flowering phase (4th month), and at fruits development stage (5th month of plantation) (Table 6).

2.1.7.4 Small Onion (Allium cepa var. aggregatum)

Trial on small onion (var. Co-ON-5) was conducted in June 2013. The seaweed bio-stimulant at 5% was sprayed as foliar application at establishment stage (10-15d), vegetative stage (25-30d), bulb formation stage (40-45d) and bulb development stage (60-65d) as shown in Table 6.

2.1.7.5 Bellary Onion (Allium cepa var. cepa)

Effect of bio-stimulant AquaSap on Bellary onion (var. Prema-178) was studied in June 2013. The application of 5% bio-stimulant was given on 10-15th (sowing establishment stage), 35-40th (vegetative stage), 60-65th (bulb formation stage) 75-80th (bulb development stage) day of sowing (Table 6).

3 Results and Discussion

All vegetable crops investigated in the present study responded well at 5% dose of bio-stimulant Aquasap (from seaweed of K. alvarezii). Highest yield was found in moringa with 52.83% over control followed by lady's finger, chillies, cabbage, garden lab lab, bellary onion, small onion, ash gourd, and snake gourd with 45.84%, 37.30%, 36.74%, 33.03%, 32.53%, 30.74%, 30.15%, and 30.02% respectively with improved quality (Table 2). Improved yield and quality in crops applied with seaweed liquid fertilizers have been well documented (Khan et al., 2009). Zodape et al. (2008) had observed that okra yielded 20.94% more over control with application of 2.5% extract of K. alvarezii and in tomato the increment when treated with 5% extract was 60.89% Zodape et al., (2011). The yield of brinjal with Eucheuma seaweed powder increased marginally higher to 41.1% (Eswaran et al., 2005). Similar kind of result with eggplant was reported when it treated with 2% extract of Ascophyllum nodosum (Bozorgi, 2012). Babu & Rengasamy (2012) observed that when chillies were treated with 1% and 2% of SLF of K. alvarezii, it increased the crop yield to 23% and 15% respectively when compare to control.

The high yield was also observed in ash gourds with 30.15% over control followed by pumpkin, snake gourd, ridge gourd, bottle gourd, bitter gourd, cucumber, watermelon and chow chow with 28.56%, 30.02%, 11.98%, 28.03%, 26.64%, 24.19%, 25.89%, and 17.34% respectively (Table 2). Ahmed & Shalaby (2012) recommend that liquid extract of *E. intestinelis* (green alga), *G. pectinutum* (red alga) or commercial seaweed liquid extract (Algreen) in addition to manure is suitable product for better vegetative growth and yield of cucumber plants. The seaweed extract of *Ascophyllum nodosum* (3g/l) applied on watermelon plant, increased the fresh weight, fruits diameter and peel thickness than control plant (Abdel-Mawgoud et al., 2010).

Higher crop yields were observed in cabbage (36.74%), cauliflower (29.61%), beetroot (28.84%), knol-khol (28.80%), radish (26.08%), potato (23.90%) and carrot (14.21%) when

compared to control plants (Table 2). The seaweed extracts powder Alga 600 and Seaforce-2 when applied on potato; it increased the dry tuber weight to 14.67% when compared with control (Sarhan, 2011). Abetz & Young (1983) observed that the yield and size of cauliflower increased when treated with *A. nodosum* extract.

The yield of treated plants in moringa, dolichos bean, bellary onion, small onion, lima bean and soybean were 52.83%, 33.03%, 32.53%, 30.74%, 25.38% and 22.10% respectively (Table 2). Soybean treated with extract of *Kappaphycus* at 15% and 12.5% showed highest grain yield of 57% and 46% respectively compared to the control and maximum straw yield was also found with treatment of 15% extract (Rathore et al., 2009). Similar kind of result in onion bulb (22.0%) when treated with *Eucheuma* seaweed powder (Eswaran et al., 2005) and high yield with improved quality of onion was found when treated with extract of *A. nodosum* (Dogra & Mandradia, 2012).

Table 2 Effect of bio-stimulant from seaweed *K. alvarezii* on yield of some vegetable crops.

Cultivar name	Variety name	Plantation type	Date of plantation	Yield increase over control (%)
Tomato	Co3 Hybrid	Seeds	03.06.12	20.94
Lady's finger	US 7902 Hybrid	Seeds	22.05.12	45.84
Brinjal	Co2 Hybrid	Seeds	03.06.12	24.53
Chillies	US 612 Hybrid	Seeds	01.01.14	37.30
Capsicum	ARKA MOHINI	Seeds	01.01.14	29.28
Ash gourd	MAH-1 Hybrid	Seeds	12.12.14	30.15
Pumpkin	ARKA CHANDAN	Seeds	12.12.14	28.56
Snake gourd	COVAI 951 F1 Hybrid	Seeds	20.07.12	30.02
Ridge gourd	US 66 Hybrid	Seeds	20.07.12	11.98
Bottle gourd	WARAD MGH-4	Seeds	20.07.12	28.03
Bitter gourd	US 475 Hybrid	Seeds	20.07.12	26.64
Cucumber	Local	Seeds	25.12.13	24.19
Watermelon	ANKUR KASHISH Hybrid	Seeds	25.12.13	25.89
Chow-chow	Green fruits	Fully matured fruits	15.08.13	17.34
Potato	KUFRI JYOTI	Seeds	15.08.13	23.90
Cabbage	MAHARANI- F1	Seeds	09.01.13	36.74
Cauliflower	SHOBHA -F1	Seeds	09.01.13	29.61
Beetroot	VALLY QUEEN	Seeds	18.01.13	28.84
Carrot	PUSA KESAR	Seeds	15.08.13	14.21
Radish	ROSHNI	Seeds	18.01.13	26.08
Lima Bean	Co2	Seeds	17.07.12	25.38
Doli Chos Bean	ANKUR GOLDY	Seeds	17.07.12	33.03
Soybean	JSS 355	Seeds	17.07.12	22.10
Moringa	PKM-1	Seeds	02.05.12	52.83
Small onion	CO-ON-5	Seeds	15.06.13	30.74
Bellary onion	PREMA - 178	Seeds	15.06.13	32.53
Knol-khol	EARLY WHITE	Seeds	18.01.13	28.80

97 Karthikeyan and Shanmugam

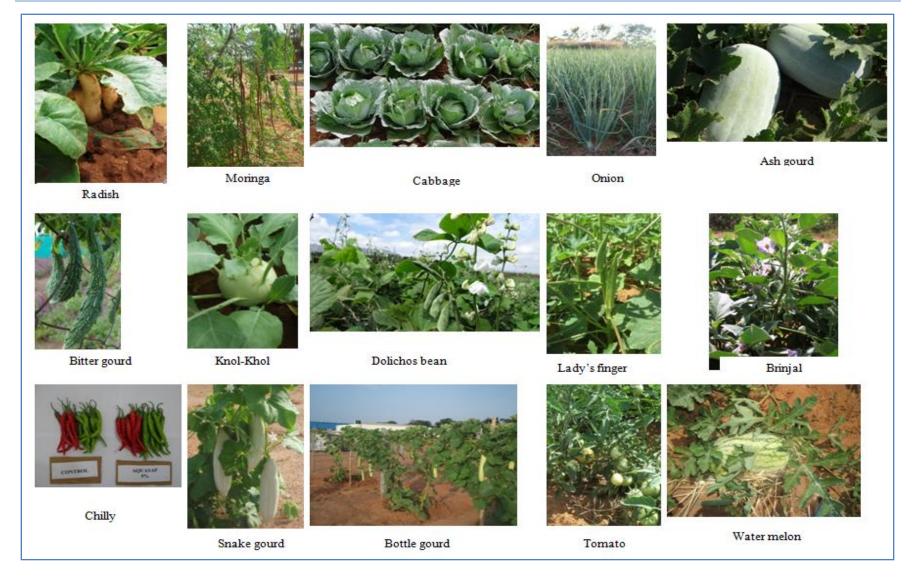


Figure 1 Some of the vegetable crops studied in the present investigation with their vegetable yield

Table 3 Dosage and application protocol of seaweed bio-stimulant used for home garden vegetable crops.

Crop name	Root dip	I Dose	II Dose	III Dose	TBS
Tomato b	Root of nurseries was dipped	10th day (After	25-30th day	45-50th day	21
	in 0.7% of bio-stimulant for 10min before transplantation.	transplantation)	(Pre-flowering stage)	(Flowering phase)	(1+5+5+5+5)
Lady's finger ^c	-	15-20th day	35-40th day	50-55th day (First	16
(Okra)		(Germination stage)	(Flowering stage)	fruits picking stage)	(1+5+5+5)
Brinjal	Root of nurseries was dipped	15-20th day	35-40th day	50-55th day (First	16
	in 0.7% of bio-stimulant for	(Germination stage)	(Flowering	fruits picking	(1+5+5+5)
	10min before transplantation.		stage)	stage)	
Chillies	Sowing	40-45th day	95-100th day	125-130th day	15
		(Vegetative stage)	(Flowering stage)	(Fruits picking stage)	(5+5+5)
	<u>Transplantation:</u> Root of nurseries was dipped in 0.7% of bio-stimulant for 10min before transplantation.	20-25th day (Days after transplantation)	60-65th day (Flowering stage)	80-85th day (First fruits picking stage)	16 (1+5+5+5)
Capsicum	Sowing	30-35th day	60-65th day	90-95th day	15
(Sweet pepper / Bell pepper)		(Vegetative stage)	(Flowering stage)	(Fruits picking stage)	(5+5+5)
	<u>Transplantation:</u> Root of nurseries is dipped in 0.7% of bio-stimulant for 10min before transplantation.	20-25th day (Days after Transplantation)	60-65th day (Flowering stage)	80-85th day (First fruits picking stage)	16 (1+5+5+5)

^a Recommended dosage of bio-stimulant: 5%; ^b IV dose at 75-80th day (Picking phase); ^c Seed treatment; Seeds were soaked for 10min in 1% of bio-stimulant before sowing, TBS - Total bio-stimulant required Per acre (L).

3.1 Effect on plant disease control

Twenty seven vegetable crops studied in the present investigation looked healthy and generally free from disease as compared to their control plants. Extract of seaweed have been reported to increase resistance of plant against pest and diseases, increase plant growth and quality yield (Jolivet et al., 1991; Verkleij, 1992; Pardee et al., 2004; Hong et al., 2007; Jeyaraj et al., 2008). Similarly, Sultana et al. (2011) had reported that number of liquid seaweed extract found to control root rotting fungi like Rhizoctonia solani, Macrophomina phaseolina, Fusarium species and root kot nematode (Meloidogyne spp.) on a variety of crops. The resistance to frost and fungal disease were reported when seaweed extract was applied to some crops (Zodape, 2001). Ara et al. (1996) had observed that extract of Sargassum spp. controlled the root rot disease in sunflower plant. Seaweed fertilizer was found to boost the resistibility adjacent to disease and in addition to reduce the insect attack (Zahid, 1999). Dogra & Mandradia (2012) had found that extract of A. nodosum significantly reduced the downy mildew severity over control in onion plant and it had also been reported that seaweed extract of Asparagopsis taxiformis found to act against phytopathogens

(Manilal et al., 2009). Lynn (1972) had observed that seaweed extract of *A. nodosum* protected *Capsicum annuum* and sweet pepper from stress to frost, microbial diseases and insect attack and increased the shelf life of fruits and better seed germination.

3.2 Effect of seed Treatment

Seed and root treatment had improved the viability of plantlets and grow vigorously as compared to control plants in the present study and it is in agreement with the literatures reports. The introductory soaking of *Triticum aestivum* seeds in 20% extracts of *Sargassum wightii* for 24 hrs gave an 11% increase in seed germination, a 63% enhance in number of lateral roots and 46% increase in shoot length in comparing to control (Kumar & Sahoo, 2011). 100% seed germination was observed in lowest concentration of SLF in black gram (Venkataraman Kumar et al., 1993) and SLF promote the seed germination as well as yield of the vegetable crops (Narasimha Rao & Reshmi Chatterjee, 2014). Treatment at 0.05% of concentrated extract of *Laminaria digitata* on *Plantago lanceolata*, *Trifolium repens* and *Avena strigosa* had given higher germination percentage (Thorsen et al., 2010).

99 Karthikeyan and Shanmugam

Table 4 Dosage and application protocol of seaweed bio-stimulant used for gourds vegetable crops.

Crop name	Seed treatment I	Dose	II Dose	III Dose	TBS
Ash gourd	Seeds were soaked for 30min in 1% of bio-stimulant	20-25th day	60-65th day (Flowering	80-85th day	16
	and incubated it for 6 days for before sowing	(Vegetative stage)	stage)	(Picking stage)	(1+5+5+5)
Pumpkin	Seeds were soaked for 30min in 1% of bio-stimulant	20-25th day	60-65th day (Flowering	80-85th day	16
	and incubate it for 6 days for before sowing	(Vegetative stage)	stage)	(Picking stage)	(1+5+5+5)
Snake gourd	Seeds were soaked for 30min in 1% of bio-stimulant	20-25th day	60-65th day (Flowering	80-85th day	16
	before sowing	(Vegetative stage)	stage)	(Picking stage)	(1+5+5+5)
Ridge gourd / Ribbed	Seeds were soaked for 30min in 1% of bio-stimulant	20-25th day	60-65th day (Flowering	80-85th day	16
gourd	before sowing	(Vegetative stage)	stage)	(Picking stage)	(1+5+5+5)
Bottle gourd	Seeds were soaked for 30min in 1% of bio-stimulant	20-25th day	60-65th day (Flowering	80-85th day	16
	before sowing	(Vegetative stage)	stage)	(Picking stage)	(1+5+5+5)
Bitter gourd	Seeds were soaked for 30min in 1% of bio-stimulant	20-25th day	60-65th day (Flowering	80-85th day	16
	before sowing	(Vegetative stage)	stage)	(Picking stage)	(1+5+5+5)
Cucumber	Seeds were soaked for 30min in 1% of bio-stimulant	10th day (Germination	35-40th day	65-70th day	16
	before sowing	phase)	(Vegetative stage)	(Flowering initiation	(1+5+5+5)
				to first picking stage)	
Watermelon	Seeds were soaked for 30min in 1% of bio-stimulant	20-25th day	60-65th day (Flowering	80-85th day	16
	before sowing	(Vegetative stage)	stage)	(Picking stage)	(1+5+5+5)

^a Recommended dosage of bio-stimulant 5%, TBS - Total bio-stimulant per acre (Lit),

Table 5 Dosage and application protocol of seaweed bio-stimulant used for cole, root and tuber vegetable crops.

Crop name	I Dose	II Dose	III Dose	TBS
Chow chow	25 - 30th day (Vegetative phase)	3rd month (Pre-flowering phase)	5th month (Flowering phase)	15 (5+5+5)
(Chayote)				
Potato b	20-25th day (Plant establishment stage)	50-55th day (Vegetative phase)	80-85th day (Early root development Phase)	20 (5+5+5+5)
Cabbage c	10-15th day (Plant establishment stage)	35-40th day (Head initiation stage)	70-75th day (Head development phase)	16 (1+5+5+5)
Cauliflower c	10-15th day (Plant establishment stage)	25-30th day (Curd initiation stage)	45-50th day (Curd development phase)	16 (1+5+5+5)
Beetroot	25-30th day (Vegetative stage)	55-60th day (Early root development stage)	80-85th day (Maturity stage)	16 (1+5+5+5)
Carrot	25-30th day (Vegetative stage)	55-60th day (Early root development stage)	80-85th day (Maturity stage)	15 (5+5+5)
Radish	10-15th day (Vegetative phase)	25-30th day (Early root development stage)	40-45th day (Maturity stage)	15 (5+5+5)
Knol-khol	10-15th day (Vegetative phase)	25-30th day (Early root development stage)	40-45th day (Maturity stage)	15 (5+5+5)

^a Recommended dosage of bio-stimulant 5%, ^b IV Dose at 100-105th day (Root maturity stage), ^c Root dip: Transplantation: Root of nurseries was dipped in 0.7% of bio-stimulant for 10min before transplantation, TBS - Total bio-stimulant per acre (Lit).

3.3 Effect of foliar application

Seaweed extract applied as foliar application found to significantly enhance the yield, growth and quality of crops (Pramanick et al., 2013). Seaweed liquid extract have gained importance to different range of crops like cereals, grasses, vegetables, species and flowers when applied through foliar application (Crouch & Van Staden, 1992). Seaweed extract is important to find out the organic sources for seed and foliar treatments for effective maintenance of vigour and viability (Dwivedi et al., 2014). The maximum yield of tomato (Zodape et al., 2011) and banana (Karthikeyan & Shanmugam, 2014) had been observed when using foliar application of *K. alvarezii* extract. Similar kind of result was observed by Pramanick et al., (2013) that the foliar application of seaweed sap improved the nutrient uptake capacity of crops.

In the present investigation, it was also observed that emerging of first flower appeared in all treated plants at least 5-10d earlier than control and similar kind of observation had been recorded in the literature. Dwivedi et al., (2014) reported that seaweed extracts not only increase the vegetative growth of the plant but it also triggers the early flowering, fruiting in crops and ultimately on seed yields. Seaweed extracts are ecologically safe, non-polluting, non-toxic, and harmless to human beings, animals and birds (Dhargalkar & Pereira, 2005). In addition to reducing the cost of inorganic fertilizers, application of seaweed bio-stimulants improves soil health, enhances the yield and quality of produce in organic vegetables production thereby increasing the domestic and international market (Chatterjee & Thirumdasu, 2014).

Conclusions

It can be concluded from the present study that 27 vegetable crops tested had responded well to bio-stimulant (Aquasap)

manufactured from seaweed *K. alvarezii*. The average yield increased from 11.98% to 45.84% with much improved vegetable quality. Therefore, the protocol used in this study will be useful to the farmers to produce organic vegetables.

Acknowledgement

The authors are very grateful to Mr. Abhiram Seth, MD, Mr. Arun Patnaik, CEO and Mr. Tanmaye Seth of AquAgri Processing Private Limited for their constant encouragement, guidance and allocation of budget to carry out the present investigations. The authors also wish to thank farmers who agreed to carry out and monitor the trials in their farm.

Conflict of interest

Authors would hereby like to declare that there is no conflict of interests that could possibly arise.

References

Abdel-Mawgoud AMR, Tantaway AS, Hafez MM, Habib HAM (2010) Seaweed extract improves growth, yield and quality of different watermelon hybrids. Research Journal of Agriculture and Biological Sciences 6: 161-168.

Abetz P, Young CL (1983) The effect of seaweed extract sprays derived from *Ascophyllum nodosum* on lettuce and cauliflower crops. Botanica Marina 26: 487-492. DOI: 10.1515/botm.1983.26.10.487.

Ara J, Ehteshamul-Haque S, Sultana V, Qasim R, Ghaffar A (1996) Effect of *Sargassum* seaweed and microbial antagonists in the control of root rot disease of sunflower. Pakistan Journal of Botany 28: 219 -223.

Table 6 Dosage and	Lapplication protoco	l of seaweed bio-	-stimulant used for	other vegetable crops.

Crop name	I Dose	II Dose	III Dose	IV Dose	TBS
Lima bean b	20-25th day	40-45th day	60-65th day (Pod	80-85th day (First	21
	(Vegetative phase)	(Flowering stage)	formation stage)	picking stage)	(1+5+5+5+5)
Dolichos beans ^b	20-25th day	40-45th day	60-65th day (Pod	80-85th day (First	21
	(Vegetative phase)	(Flowering stage)	formation stage)	picking stage)	(1+5+5+5+5)
Soybean b	20-25th day	40-45th day	60-65th day (Pod	80-85th day (First	21
	(Vegetative phase)	(Flowering stage)	formation stage)	picking stage)	(1+5+5+5+5)
Moringa ^b	25-30th day	3rd month (Pre-	4th month	5th month (Fruits	21
	(Nurseries stage)	flowering phase)	(Flowering phase)	development stage)	(1+5+5+5+5)
Small onion	10-15th day (Sowing	25-30th day	40-45th day (Bulb	60-65th day (Bulb	20
	to establishment	(Vegetable stage)	formation stage)	development stage)	(5+5+5+5)
	stage)				
Bellary onion	10-15th day (Sowing	35-40th day	60-65th day (Bulb	75-80th day (Bulb	20
	to establishment	(Vegetable stage)	formation stage)	development stage)	(5+5+5+5)
	stage)				

^a Recommended dosage of bio-stimulant 5%, ^b Seed treatment - Seeds were soaked for 10min in 1% of bio-stimulant before sowing, TBS

⁻ Total bio-stimulant per acre (Lit).

Ahmed YM, Shalaby EA (2012) Effect of different seaweed extracts and compost on vegetative growth, yield and fruit quality of cucumber. Journal of Horticultural Science & Ornamental Plants 4 : 235-240. DOI: 10.5829/idosi.jhsop.2012.4.3.252.

Anisimov MM, Chaikina EL (2014) Effect of seaweed extracts on the growth of seedling roots of soybean (*Glycine max* (L.) Merr.) seasonal chages in the activity. International Journal of Current Research and Academic Review 2: 19-23.

Babu S, Rengasamy R (2012) Effect of *Kappaphycus alvarezii* SLF treatment on seed germination, growth and development of seedling in some crop plants. Journal of Academia and Industrial Research 1: 186-95.

Bozorgi HR (2012) Effects of foliar spraying with marine plant *Ascophyllum nodosum* extract and nano iron chelate fertilizer on fruits yield and several attributes of egg plant (*Solanum melongena* L.). ARPN Journal of Agricultural and Biological Science 7: 357-362.

Chatterjee R, Thirumdasu RK (2014) Nutrient management in organic vegetable production. International Journal of Food, Agriculture and Veterinary Sciences 4: 156-170.

Crouch IJ, Van Staden J (1992) Effect of seaweed concentrate on the establishment and yield of greenhouse tomato plants. Journal of Applied Physiology 4: 291-296. DOI: 10.1007/BF02185785.

Dhargalkar VK, Pereira N (2005) Seaweed: Promising plant of the millennium.Science and Culture 71: 60-66 available on http://drs.nio.org/drs/handle/2264/489 access on 14th November, 2015.

Dogra BS, Mandradia RK (2012) Effect of seaweed extract on growth and yield of onion. International Journal of Farm Sciences 2: 59-64.

Dwivedi SK, Meshram MR, Pal A, Pandey N, Ghosh A (2014) Impact of natural organic fertilizer (seaweed sap) on productivity and nutrient status of black gram (*Phaseolus mungo* L.). The Bioscan 9: 1535-1539.

Eswaran K, Ghosh PK, Siddanta AK, Patolia JS, Periyasamy C, Mehta AS, Mody KH, Ramavat BK, Prasad K, Rajyaguru MR, Reddy SKCR, Pandya JBP, Tewari A (2005) Integrated method for production of carrageenan and liquid fertilizer from fresh seaweeds. US patent 6893479 B2, field August 19, 2002. http://www.google.co.in/patents/US6893479

Hong DD, Hien HM, Son PN (2007) Seaweeds from Vietnam used for functional food, medicine and bioferilizer. Journal of Applied Phycology 19: 817-826. DOI: 10.1007/s10811-007-9228-x.

Jayaraj J, Wan A, Rahman M, Punja ZK (2008) Seaweed extracts reduces foliar fungal disease on carrot. Crop Production 27: 1360-1366. DOI:10.1016/j.cropro.2008.05.005.

Jolivet E, Langlais JI, De JF, Morot-Gaudry JI, Langais DE (1991) Extracts of marine algae: Phytoactive properties and agronomic value. Annee Biologique 30: 109-126.

Karthikeyan K, Shanmugam M (2014) Enhanced yield and quality in some banana varieties applied with commercially manufactured bio-stimulant AquaSap from sea plant *Kappaphycus alvarezii*. Journal of Agricultural Science and Technology B 4: 621-631. DOI: 10.17265/2161-6264/2014.08.004.

Khan W, Rajirath UP, Subramanian S, Jithesh MN, Rayorath P, Hodges DM, Critchley AT, Craige JS, Norrie J, Prithiviraj B (2009) Seaweed extract as biostimulants of plant growth and development. Journal of Plant Growth Regulator 28: 386-399. DOI 10.1007/s00344-009-9103-x.

Kumar G, Sahoo D (2011) Effect of seaweed liquid extract on growth and yield of *Triticum aestivum* var. Pusa Gold. Journal of Applied Phycology 23: 251-255. DOI: 10.1007/s10811-011-9660-9.

Lynn LB (1972) The Chelating Properties of Seaweed Extract, *Ascophyllum nodosum* vs. Macrocystis perifera on the Mineral Nutrition of Sweet Peppers, *Capsicum annuum*. Clemson University, Clemson, S.C.

Manilal A, Sujith S, Kiran GS, Selvin J, Shakir C, Gandhimathi R, Lipton AP (2009) Antimicrobial potential and seasonality of red algae collected from the southwest coast of India tested against shrimp, human and phytopathogens. Annals of Microbiology 59: 207-219. DOI: 10.1007/BF03178319.

Narasimha Rao GM, Reshmi Chatterjee (2014) Effect of seaweed liquid fertilizer from *Gracilaria textorii* and *Hypnea musciformis* on seed germination and productivity of some vegetable crops. Universal Journal of Plant Science 2: 115-120. DOI: 10.13189/ujps.2014.020701.

Pise NM, Sabale AB (2010) Effect of seaweed concentrates on the growth and biochemical constituents of *Trigonella foenum-graecum* L. Journal of Phycology 2: 50-56.

Pardee KI, Ellis P, Bouthillier M, Towers GHN, French CJ (2004) Plant virus inhibitors from marine algae. Canadian Journal of Botany 82: 304-309. DOI: 10.1139/b04-002.

Pramanick B, Brahmachari K, Ghosh A (2013) Effect of seaweed saps on growth and yield improvement of green gram. African Journal of Agriculture Research 8: 1180-1186. DOI: 10.5897/AJAR12.1894.

Pramanick B, Brahmachari K, Ghosh A, Zodape ST (2014) Foliar nutrient management through *Kappaphycus* and *Gracilaria* saps in rice-potato-green gram crop sequence. Journal of Scientific & Industrial Research 73: 613-617.

Prasad K, Das AK, Oza MD, Brahmbhatt H, Siddhanta AK, Meena R, Eswaran K, Rajyaguru MR, Ghosh PK (2010) Detection and quantification of some plant growth regulators in a seaweed-based foliar spray employing a mass spectrometric technique sans chromatographic sepatation. Journal of Agricultural and Food Chemistry 58: 4594-4601. DOI: 10.1021/jf904500e.

Rathore SS, Chaudhary DR, Boricha GN, Ghosh A, Bhatt BP, Zodape ST, Patolia JS (2009) Effect of seaweed extract on the growth, yield and nutrient uptake of soybeen (*Glycine max*) under rainfed conditions. South African Journal of Botany 75: 351-355. doi:10.1016/j.sajb.2008.10.009.

Sarhan TZ (2011) Effect of humic acid and seaweed extracts on growth and yield of potato plant (*Solanum tuberosum* L) DESIREE CV. Mesopotamia Journal of Agriculture 39: 19-27.

Sridhar S, Rengasamy R (2010) Significance of seaweed liquid fertilizers for minimizing chemical fertilizers and improving yield of *Arachis hypogaea* under field trial. Recent Research in Science and Technology 2: 73-80.

Sumkiman, Faturrahman, Imy Suci Rohyani, Hilman Ahyadi (2014) Short communication: Growth of seaweed *Eucheuma cottonii* in multi trophic sea farming systems at Gerupuk Bay, Central Lombok, Indonesia. Bioscience 6: 82-85. DOI: 10.13057/nusbiosci/n060113.

Sultana V, Baloch GN, Ara J, Ehteshamul-Haque S, Tariq RM, Athar M (2011) Seaweed as an alternative to chemical pesticide for the management of root diseases of sunflower and tomato. Journal of Applied Botany and Food Quality 84:162-168.

Thorsen MK, Woodward S, McKenzi BM (2010) Kelp (*Laminaria digitata*) increases germination and affects rooting and plant vigour in crops and native plants from an arable

grassland in the Outer Hebrides, Scotland. Journal Coastal Conservation 14: 239-247. DOI:10.1007/s11852-010-0091-6.

Verkleij FN (1992) Seaweed extracts in agriculture and horticulture: A review. Biological . Biological Agriculture & Horticulture: An International Journal for Sustainable Production Systems 8:309-326.

Vanitha SM, Chaurasia SNS, Singh PM, Prakash S Naik (2013) Vegetable Statistics. Technical Bulletin No. 51:001-250 available on www.iivr.org.in/.../Technical%20Bulletins/7.%20Vegetable%2 0Statistics.pdf access on 14th November, 2015.

Venkataraman Kumar V, Mohan VR, Murugeswari R, Muthusamy M (1993) Effect of crude and commercial seaweed extracts on seed germination and seedling growth in green gram and black gram. Seaweed Research and Utilization 16: 23-27.

Zahid PB (1999) Preparation of organic fertilizer from seaweed and its effect on the growth of some vegetables and ornamental plants. Pakistan Journal of Biological Science 2:1247-1277.

Zodape ST (2001) Seaweeds as a Biofertilizer. Journal of Scientific & Industrial Research 60: 378-382.

Zodape ST (2009) Effect of *Kappaphycus alvarezii* (Doty) Doty ex silva. extract on grain quality, yield and some yield components of wheat (*Triticum aestivum* L.). International Journal of Plant Production 3: 97-10.

Zodape ST, Gupta A, Bhandari SC, Rawat US, Chaudhary DR, Eswaran K, Chikara J (2011) Foliar application of seaweed sap as biostimulant for enhancement of yield and quality of tomato (*Lycopersicon esculentum* Mill.). Journal of Scientific & Industrial Research 70: 215 - 219.

Zodape ST, Kawarkhe VJ, Patolia JS, Warade AD (2008) Effect of liquid seaweed fertilizer on yield and quality of Okra (*Abelmoschus esculentus L*). Journal of Scientific & Industrial Research 67:1115-1117.